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TITLE: OPTICAL METHODS AND SYSTEMS FOR TISSUE ANALYSIS

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Optical Methods and Systems for Tissue Analysis

TECHNICAL FIELD

The invention relates to tissue analysis, and more particularly to characterizing tissue by analyzing speckle patterns formed by light reflected from tissue.

5

BACKGROUND

"Speckle" is an interference phenomenon that occurs when coherent light (e.g., laser light) is reflected from a rough or multiply scattering sample onto a detection plane. Due to scattering of photons from and within the sample, different photons travel different distances to the detection plane. As a result, the light reflected or backscattered from the sample, if
10 spatially and temporally coherent, interferes at the detection plane, producing a grainy pattern known as "speckle."

Researchers have used speckle pattern analysis to study dynamic movement of tissue *in vivo*. For example, speckle has been used to measure vibrations of tissue, V. Tuchin et al., "Speckle interferometry in the measurements of biotissues vibrations," SPIE, 1647: 125
15 (1992), and to measure strain in vascular and cortical tissue in response to forced movement of the tissue, Sean J. Kirkpatrick et al., "Laser Speckle Microstrain Measurement in Vascular Tissue," SPIE, 3598: 121-128 (1999); and Sean J. Kirkpatrick and Brent W. Brooks, "Micromechanical Behavior of Cortical Bone as Inferred from Laser Speckle Data," J. Biomedical Materials Research, 39(3): 373-79 (1998). Researchers have also used speckle to
20 study blood flow and lymph flow. B. Ruth, "Blood Flow Determination by the Laser Speckle Method," Int'l J. Microcirc: Clinical and Experimental, 9(1): 21-45 (1990); and A.A. Bednov et al., "Investigation of Statistical Properties of Lymph Flow Dynamics Using Speckle-Microscopy," SPIE, 2981: 181-90 (1997).

SUMMARY

25 The invention is based on the discovery that tissues can be analyzed *in vivo* using laser speckle to measure Brownian motion.

In general, the invention features a method of analyzing tissue, e.g., *in vivo*, by illuminating a tissue with coherent light, such as laser light; receiving light reflected from the tissue at a detector to form a series of speckle patterns; and analyzing changes in the speckle

patterns at time intervals sufficient to measure changes caused by Brownian motion of objects within the tissue. For example, the speckle pattern can be a far field image formed at the detector. The method can further include compensating for extrinsic motion, such as a heartbeat, to isolate the Brownian motion. In this method, the illuminating step can include
5 providing an invasive device coupled to a light source, passing the device into a patient, placing the device in proximity to the tissue, and shining coherent light from the light source onto the tissue.

The invasive device can be, e.g., a catheter, an endoscope, or a laparoscope. The device can be placed in direct contact with the tissue. The device can include a catheter
10 having a first fiber that transmits light from the light source to the tissue, and a fiber array that receives light remitted from the tissue. The analyzing step can include comparing each of the series of speckle patterns to a reference speckle pattern, and quantifying the differences between each pattern and the reference pattern. For example, the analyzing step can include digitizing each of the speckle patterns, and the quantifying step can include evaluating a
15 maximum cross-correlation between each pattern and the reference pattern. The analyzing step can further include determining a decorrelation rate for the speckle patterns, or analyzing spatial characteristics of the speckle pattern to deduce structural characteristics of the tissue.

In variations, the method can include illuminating multiple sections of the tissue in succession, forming a separate series of speckle patterns for each respective section of the
20 tissue, and then analyzing each separate series of speckle patterns and comparing the separate series to deduce structural differences between the respective sections of the tissue.

In certain embodiments, the method includes gathering reflected light at a light receptor and transmitting the gathered light to the detector, and compensating for extrinsic motion by coupling the receptor to the tissue. Compensating for extrinsic motion can also be
25 done by excluding changes in the speckle patterns caused by non-random motion during the analysis step. Extrinsic motion can also result, for example, from blood flowing between the tissue and the reflector. In those cases, the compensating step can include replacing the blood with a transparent solution.

In another embodiment, the invention features a method of determining the
30 susceptibility to rupture of an atherosclerotic plaque having a lipid pool and a fibrous cap. The method includes illuminating the plaque with coherent light; receiving light reflected

from the plaque at a detector to form a series of speckle patterns; gathering speckle pattern data at time intervals sufficient to measure Brownian motion within the lipid pool; and assessing the plaque's vulnerability to rupture from the amount of Brownian motion.

5 The method can further include analyzing spatial characteristics of the speckle pattern data to determine structural characteristics of the plaque, for example, by assessing the thickness of the fibrous cap. In that case, a plaque is considered vulnerable to rupture if the thickness of the fibrous cap is less than about 60 microns. The method can also be used to assess the viscosity of the lipid pool, wherein the plaque is considered vulnerable to rupture if the viscosity of the lipid pool has a time constant of less than about 200 milliseconds, and
10 considered likely to rupture if the viscosity of the lipid pool has a time constant of less than about 100 milliseconds.

The invention also includes a method of detecting a vulnerable atherosclerotic plaque having a lipid pool and a fibrous cap within a blood vessel by illuminating a segment of the blood vessel in vivo with coherent light; receiving light reflected from the interior vessel wall
15 of the segment at a detector to form a series of speckle patterns; gathering speckle pattern data at time intervals sufficient to measure Brownian motion within the interior vessel wall; and comparing the speckle pattern data to a known speckle pattern for a normal blood vessel and a known speckle pattern for an atherosclerotic plaque; wherein speckle pattern data corresponding to a speckle pattern for an atherosclerotic plaque indicates the segment of the blood vessel contains an atherosclerotic plaque. In this method, spatial characteristics of the
20 speckle pattern data can be analyzed to determine structural characteristics of the plaque as described herein.

In another aspect, the invention features a fiber optic probe for detecting speckle patterns in a sample. The probe includes a catheter including a rotatable inner shaft and a
25 transparent outer sheath; a fiber array housed within the shaft and comprising a central optical fiber for transmitting incident light to the sample and multiple optical fibers for transmitting light remitted from the sample; and a mirror arranged near a distal end of the shaft to reflect light passing through the fiber array onto a sample outside the transparent outer sheath and back from the sample through the fiber array. In some embodiments, the
30 shaft can rotate 360 degrees within the sheath, and an inflatable balloon can be connected to the sheath.

The invention further includes an optical system for detecting speckle patterns in a sample. The system has a fiber optic probe as described herein; a coherent light source connected to the central optical fiber within the fiber array; a detector to receive light remitted from the sample; and a processor to process the remitted light and to analyze
5 speckle patterns remitted from the sample. For example, the processor can include a reference speckle pattern or a whole library of reference speckle patterns, e.g., for healthy and diseased tissue. The system can also include an analog-digital converter to convert the analog remitted light into a digital signal.

As used herein, "tissue" means any biological structure in or on a body. Tissue
10 includes aggregates of cells, growths, and deposits such as plaque that may contain lipids or other components. Specific components of plaques that can be investigated include lipid pools, calcifications, fibrous regions, and fibrous caps.

"Speckle" is an interference phenomenon that occurs when coherent light is reflected from a rough or multiply scattering sample onto a detection plane. A "speckle pattern" is the
15 intensity pattern that results from speckle interference.

"Brownian motion" is the random motion of cells, molecules, and other subcomponents within tissue.

"Extrinsic motion" is motion other than Brownian motion. Extrinsic motion includes gross, macroscopic motion of tissue, such motion caused by a heartbeat or other muscle
20 motion, and the movement of fluids in the body, such as blood flow.

Unless otherwise defined, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs. Although methods and materials similar or equivalent to those described herein can be used in the practice or testing of the present invention, suitable methods and materials are
25 described below. All publications, patent applications, patents, and other references mentioned herein are incorporated by reference in their entirety. In case of conflict, the present specification, including definitions, will control. In addition, the materials, methods, and examples are illustrative only and are not intended to be limiting.

Other features and advantages of the invention will be apparent from the following
30 detailed description and from the claims.

BRIEF DESCRIPTION OF THE DRAWINGS

Fig. 1 is a cross-sectional schematic of an optical catheter for gathering speckle data from tissue *in vivo*.

Fig. 2A is a cross-sectional schematic illustrating the catheter of Fig. 1, with an attached angioplasty balloon, inserted within a blood vessel, with the balloon deflated.

Fig. 2B is a cross-sectional schematic illustrating the catheter of Fig. 1, with an attached angioplasty balloon, inserted within a blood vessel, with the balloon inflated.

Fig. 3 is a speckle pattern produced from a cadaveric human aorta using incident light of $\lambda = 632.8$ nm.

Fig. 4 is a schematic illustrating reflectance of incident light from an atherosclerotic plaque.

Fig. 5 is representative raw data speckle images, edge images, and cross-correlation images used to assess the viscosity of a lipid-rich, atherosclerotic plaque in a human aorta.

Fig. 6 is representative raw data speckle images, edge images, and cross-correlation images used to assess the viscosity of normal human aorta tissue.

Fig. 7 is an exponential graph showing speckle decorrelation of an atherosclerotic plaque and aortic tissue over a time interval.

DETAILED DESCRIPTION

At a microscopic level, most tissue is not static. Individual cells move within intercellular fluids, cellular organelles move within cells, and large molecules move back and forth between cells. In non-cellular tissue deposits such as plaques, components such as proteins, lipids, and other molecules also exhibit local motion. These local motions, called "Brownian motion," are essentially random in nature. Measuring and characterizing the Brownian motion of tissues can provide useful information about the structure, composition, and stability of the tissue.

The invention relates to using laser speckle to measure Brownian motion of tissue *in vivo* to gather information about the tissue. In general, coherent light is reflected from a tissue to form a speckle pattern at a detector. Due to motion of reflectors in the tissue, the speckle pattern changes over time, or "decorellates." By monitoring the rate of decorrelation,

while compensating for "extrinsic," or non-Brownian motion of the tissue, Brownian motion in the tissue can be isolated and measured.

In some embodiments of the invention, speckle analysis is used to measure Brownian motion in atherosclerotic plaques to detect plaques that are vulnerable to rupture, and, more specifically, to determine the plaque's vulnerability to rupture. In these embodiments, a modified optical catheter (probe) or other instrument is inserted into a blood vessel (e.g., artery) to locate these plaques, and once a plaque is located, the probe is moved into the proximity of the specific atherosclerotic plaque. Light reflected from the interior wall of the blood vessels, and/or from a plaque, is collected and transmitted to a detector, where a speckle pattern is formed. The speckle patterns of normal tissue and plaque tissue (especially vulnerable plaque tissue) are different, and these differences can be used to detect the plaques. Thereafter, e.g., while compensating for extrinsic motion of the plaque, the speckle pattern is monitored over time to calculate the pattern's rate of decorrelation. From this decorrelation rate, the degree of Brownian motion in the plaque, and therefore the plaque's vulnerability to rupture, can be assessed.

I. Atherosclerotic Plaques

Rupture of an atherosclerotic plaque can lead to acute myocardial infarction, which is a leading cause of death in industrialized countries. When an atherosclerotic plaque ruptures, lipids from the plaque enter the vessel lumen, potentially causing thrombosis, arterial occlusion, myocardial ischemia, and infarction.

According to recent research, plaques vulnerable to rupture generally have a thin, unstable, fibrous cap and a compliant, or "viscous," lipid pool. See, e.g., Virmani et al., "Lesions from sudden coronary death: A comprehensive morphological classification scheme for atherosclerotic lesions," *Arterioscler. Thromb. Vasc. Bio.*, 20:1262-75 (2000) and Lee et al., "The Unstable Atheroma," *Arteriosclerosis, Thrombosis & Vascular Biology*, 17:1859-67 (1997). The viscous lipid pool applies force to the fibrous cap, compromising the cap and causing rupture. The greater the Brownian motion in the lipid pool, the greater the "viscosity" of the pool, and the more likely the plaque will rupture. Assessing Brownian motion in the lipid pool and measuring the thickness of the fibrous cap *in vivo*, therefore, helps to identify plaques likely to rupture, allowing intervention.

II. Speckle Image Formation

Referring to Fig. 1, a specially modified optical catheter 10 includes a rotatable inner shaft 12 and a transparent outer sheath 14. The inner shaft 12 houses a fiber array 15 and a mirror 16 near its distal end 18. A central fiber 20 in the fiber array connects to a fixed
5 optical fiber 21 that extends from the catheter proximally to a light source 22.

In operation, coherent light, such as laser light, from light source 22 is transmitted via beam-splitter 22a, through the fixed optical fiber 21 and central fiber 20 and onto center 23 of mirror 16. From mirror 16, the light is reflected to a tissue sample 24, such as an atherosclerotic plaque. Outer sheath 14 can be placed directly in contact with sample 24, or
10 can be positioned a short distance, e.g., 1 mm to 10 cm away from the sample. Light enters sample 24, where it is reflected by molecules, cellular debris, proteins, compounds (e.g., cholesterol crystals), and cellular microstructures (such as organelles, microtubules) within the sample. Light remitted from the sample (arrows 26) reflects from mirror 16 to the fibers of array 15, and is then transmitted by array 15 to a planar charge-coupled device (CCD), or a
15 linear or two-dimensional) detector 22b, via a beam-splitter 22a, e.g., located within light source 22. Due to interference, a speckle pattern forms at the CCD detector. The resulting speckle pattern is then digitized by an analog-digital converter, and analyzed using the procedures described in the analysis section below.

The entire shaft 12 can rotate 360 degrees in the direction of arrow R, allowing
20 catheter 10 to gather images around the entire circumference of a sample. For example, catheter 10 can gather images of a plaque around the circumference of a vessel wall.

Since only a few fibers are required to gather adequate speckle data, the diameter of the catheter can be less than 500 μm . Larger diameters are also possible.

Many other types of instruments can be used to gather speckle data. For example, the
25 optics of catheter 10 can be integrated into other types of instruments, such as endoscopes or laparoscopes. The optics can also form a stand-alone unit passed into the accessory port of standard endoscopes or laparoscopes, or integrated into another type of catheter, such as dual-purpose intravascular ultrasound catheter.

The optics can also include a lens that focuses the remitted light 26 onto the distal
30 ends of the fibers in array 15. The lens would allow formation of a "near field image" (near the sample sight) rather than a "far field image" (at the detector).

The catheter can include a polarization filter to remove all but a certain type of polarized light. For example, a cross-polarized filter would allow only light having a polarization perpendicular to the incident light to reach the detector, while a parallel polarized filter would allow only light having the same polarization as the incident light to pass. Since multiply scattered light is less likely to retain its initial polarization than single scattered light, polarization filters can be used to bias the data toward multiply scattered or single scattered light. Such bias can be used to deduce information about the structure of the sample, since light which has penetrated deeper into the sample will be more highly scattered than light reflected from the surface or remitted from near the surface.

Instead of a CCD, the detector can be, e.g., a photographic plate or an array of photodetectors. The light source can illuminate the sample with continuous light or synchronized pulses.

Rather than transmitting the light to the sample through optical fibers, it is also possible to shine light onto a sample in free space. For example, in an open surgical procedure, coherent light in free space could be directed onto a sample with mirrors, and the remitted light then directed to a fiber array. In such free space embodiments, the light source can be, e.g., as far as one meter, or more, away from the sample.

III. Isolation of Brownian Motion

To simplify determining the viscosity of a plaque's lipid pool from changes in a speckle pattern, the temporal changes in the pattern should indicate movement of reflectors within the plaque, but not indicate movement of the plaque itself or movement of reflectors between the detector and the plaque. In other words, the changes in the plaque's speckle pattern preferably reflect Brownian motion, but not extrinsic motion.

To isolate Brownian motion, data is gathered: (1) at time intervals sufficient to detect Brownian motion; and (2) in a manner that compensates for extrinsic motion.

For a time interval to be sufficient to detect Brownian motion, the interval must be long enough to allow for movement of reflectors in the lipid pool, but short enough that the random Brownian movements do not cancel out. For atherosclerotic plaque, an appropriate time interval is about 1-200 ms. Shorter time periods may also be possible. If the time intervals are longer, then changes in the speckle pattern may not adequately differentiate

rapid Brownian movement (indicating high viscosity) from slower Brownian movement (indicating low viscosity).

In the atherosclerotic plaque example, the two most likely sources of extrinsic motion are gross movement of the vessel lumen and plaque tissue due to heartbeats, and blood flow
5 between the plaque and the catheter.

To compensate for gross motion of the plaque due to heartbeats, at least two alternatives are possible. First, the fiber array 15 can be coupled to the plaque tissue using, e.g., an angioplasty balloon. Referring to Fig. 2A, in one embodiment, a balloon 28 is attached to outer sheath 14, on a far side 30 of the catheter. Once the catheter is positioned
10 within a blood vessel in proximity to the plaque, the balloon is inflated. Referring to Fig. 2B, the inflated balloon abuts the vessel wall 32, and presses the catheter against plaque 24, such that a distal region of outer shaft 14 is in direct contact with the plaque. With the catheter coupled to plaque 24 as shown in Fig. 2B, fiber array 15 will move with the plaque when the heart beats, and the gross motion of the plaque will not significantly affect the speckle
15 pattern.

Other methods of coupling the catheter to the plaque are also possible. For example, instead of placing the balloon to the side of the catheter, the balloon can surround the catheter. In this arrangement, a transparent balloon surrounds outer sheath 14, but is also attached to the sheath. When the balloon is inflated, the balloon is squeezed between plaque
20 24 and wall 32 of the vessel. The balloon, therefore will be in direct contact with the plaque, and will move with the plaque when the heart beats. Since the balloon is attached to shaft 14, and shaft 14 is coupled to array 15, movement of the vessel wall will not significantly affect the speckle pattern. Additional methods of coupling the catheter to tissue can also be used, including methods that do not employ an angioplasty balloon.

25 A second method of compensating for movement caused by heartbeats is to gather data between heartbeats. In this method, data is gathered during the relatively still PR interval of the diastole of the heartbeat (when the left ventricle is filling with blood). The PR interval lasts for about 0.12-0.2 seconds, providing sufficient time to detect Brownian motion. To insure that data is gathered during diastole, the timing can be computer-
30 controlled or the detector can be linked to an ECG signal, and programmed to gather data only during the PR interval.

To compensate for blood flow between the catheter and the plaque, the catheter can be placed in direct contact with the plaque tissue, as described above, thereby preventing blood from flowing between the detector and the plaque. Alternatively, blood flowing between the plaque and the catheter can be removed and replaced with clear saline solution.

5 Finally, rather than compensating for extrinsic motion while gathering data, one can compensate for extrinsic motion during the analysis phase by mathematically excluding the extrinsic motion from the analysis, as described below.

IV. Analysis of Speckle Data

10 Fig. 3 illustrates a typical speckle pattern 40 formed by reflecting light from the wall of a healthy blood vessel. X and Y coordinates overlay pattern 40 to facilitate mathematical description of the pattern. The pattern includes dark patches, where destructive interference dominates, and brighter patches, where constructive interference (or no interference) dominates. Very subtle movements of reflectors within the multiply scattering sample alter
15 the speckle pattern.

By analyzing a series of speckle patterns formed from light reflected from a plaque, one can estimate: (a) the viscosity of a plaque's lipid pool; and (b) the thickness of plaque's fibrous cap. From either or both of these types of data, the plaque's vulnerability to rupture can be assessed.

A. Determining Viscosity of a Plaque's Lipid Pool

20 There are a number of methods of analyzing speckle data to determine the viscosity of a plaque's lipid pool. By way of example, one method is described in detail in this section and in the Example section below. This method includes: (1) gathering a series of speckle
25 images at short, discrete time intervals; (2) eliminating diffuse reflectance from the data; (3) creating cross-correlation images comparing the speckle images in the series; (4) calculating the maximum correlation between each pair of images to create a one-dimensional data set over time; (5) calculating the rate of decorrelation from the data set; and (6) from the rate of decorrelation, assessing the plaque's viscosity and vulnerability to rupture.

30 First, using the detection system described above, a series of speckle images are gathered for a plaque at discrete intervals over a period of time. For example, speckle images

can be gathered, e.g., at intervals of every 1, 5, 10, 20, or 30 ms for a time period of, e.g., 200 ms. In general, the shorter the time intervals, the shorter the time period over which data can be gathered. For longer time intervals, such as 30 ms, data can be gathered for, e.g., 1-2 seconds.

5 Second, to isolate the speckle pattern, the background, non-coherent diffuse reflectance is eliminated from the images. A number of techniques can be used to eliminate the tissue's diffuse reflectance. For example, the raw data speckle images can be converted to edge images. Edge images are spatial derivatives of the raw data images; an edge image reflects the change in intensity of an image as a function of space, at all points in the image,
10 rather than the intensity itself. Known techniques of edge detection include convolution of the image by a kernel (e.g., Sobel or Robert), Morph gradient (subtraction of an eroded, dilated, closed, or opened image by its original), or high pass filtering. Other methods of eliminating background diffuse reflectance include homomorphic filtering, local histogram equalization, or using an optical setup with a small aperture. All of these techniques are well
15 known, and are described, e.g., in Gonzalez, R.C. and Wintz, P., "Digital Image Processing" (Addison-Wesley Publishing Company, Reading MA, 1987) and Jain, Anil, K., "Fundamentals of Digital Image Processing" (Prentice Hall, Englewood Cliffs, NJ, 1987).

 After eliminating the non-coherent background reflectance, each speckle image (or edge image) is compared to a reference image in the series (e.g., the $t = 0$ image) to create a
20 series of cross-correlation images. The cross-correlation images reflect the degree of correlation between the two images as a function of space. From each cross-correlation image, the maximum correlation peak (i.e., the amount of correlation at the point of maximum correlation) is determined using the equation:

25
$$g(t) = \max \left[\iint I(x, y, 0) I(x + x', y + y', t) dx' dy' \right] \quad (1)$$

 where $g(t)$ is the cross-correlation function, $I(x, y)$ is intensity of the interference at a point (x, y) in the pattern, and t is time. Two-dimensional cross-correlation functions are described generally in Jae S. Lim, "Two-Dimensional Signal Processing" (Prentice Hall, Englewood
30 Cliffs, NJ, 1990) and Jain, Anil, K., "Fundamentals of Digital Image Processing" (Prentice Hall, Englewood Cliffs, NJ, 1987).

By performing the maximum correlation calculations, the cross-correlation images are reduced to a one-dimensional data set as a function of time (i.e., a series of correlation values, each value associated with a time t). From this series of correlation values, a time constant, τ , is calculated, where τ represents the rate of decorrelation. The time constant is
5 the amount of time it takes $g(t)$ to reach $(1/e)g(0)$.

The max function of equation (1) is not the only possible mechanism for reducing the cross-correlation images to a number. For example, image comparisons can be reduced to a representative value by evaluating the cross-correlation function:

$$g(x, y, x_0, y_0, t) = \iint w(x' - x_0, y' - y_0) I(x, y, 0) I(x + x', y + y', t) dx' dy' \quad (2)$$

at a point, such as $x = y = 0$. However, using a point to reduce the image to a value, rather than a max function, would not compensate for the "memory effect" of first order correlation of speckle patterns in turbid media. This "memory effect" is described in Feng et al., Science
15 251: 633-39 (1991). Advantages to using a point are that a minimum number of fibers and detectors can be used.

From the rate of decorrelation, represented by time constant τ , the viscosity of the plaque's lipid pool can be assessed. In general, the larger τ , the lower the Brownian motion in the lipid pool, and the greater the pool's viscosity. On the other hand, the smaller τ , the
20 greater the Brownian motion in the lipid pool, and the lower the viscosity. The lower the viscosity, the more stresses are exerted on the cap, making the plaque more vulnerable

This information, the viscosity of the plaque's lipid pool, can be used to identify plaques likely or vulnerable to rupture. Specifically, if τ is about 40-100 ms or lower, then the plaque is likely to rupture, and intervention is warranted. If the plaque τ is about 100-200
25 ms, then the plaque is somewhat vulnerable, but not yet likely to rupture, and should be monitored over time. If the plaque τ is about 200-300 ms, then the plaque is not vulnerable. Non-plaque covered, healthy vessel wall generally has a time constant greater than 300 ms.

B. Determining Thickness of a Plaque's Fibrous Cap

30 In addition to determining the viscosity of the lipid pool in the plaque, the speckle data can be analyzed to deduce spatial characteristics of the plaque, including the thickness of the fibrous cap. As discussed above, a thin fibrous cap is another indication that a plaque

is vulnerable to rupture. The combination of data relating to viscosity and cap thickness provides the most accurate assessment of plaque vulnerability, although the two characteristics can be assessed and analyzed independently.

Referring to Fig. 4, a typical plaque 50 includes a fibrous cap 52 and a lipid pool 54.

- 5 Photons that enter plaque 50 (arrow A) are internally scattered by reflectors within the plaque, such as collagen in fibrous cap 52 and lipids in pool 54. The various photons, therefore, exit the plaque at different locations (arrows B). As a result, the speckle pattern (see Fig. 2) has a diameter considerably larger than the width of the original light beam.

- The thickness of fibrous cap 52 can be deduced by comparing different regions of the resulting speckle pattern. Referring again to Fig. 3, light forming intensity signals in the
10 outer portion 60 of the pattern traveled greater distances than light forming signals near the center 62 of the pattern. Thus, outer portion 60 of the pattern is formed by photons that, on average, penetrated deeper into the plaque than photons forming center 62. By calculating
15 separate time constants for separate regions of the speckle pattern, the viscosity of the plaque at different depths can be determined. Since the fibrous cap generally exhibits less Brownian motion than the lipid pool, the thickness of the fibrous cap can be estimated from spatially dependent data.

- To estimate the thickness of the fibrous cap, separate max cross-correlation functions are described for separate, small regions of the pattern. Each region is defined by a window,
20 w , centered at (x_0, y_0) :

$$g(x_0, y_0, t) = \max \left[\iint w(x' - x_0, y' - y_0) I(x, y, 0) I(x + x', y + y', t) dx' dy' \right] \quad (3)$$

- Time constants are then calculated from the cross-correlation data for each window,
25 in the manner described above. The variation of τ as a function of the distance from the center of the speckle pattern (i.e., as a function of $(x^2 + y^2)^{1/2}$) can then be analyzed to determine the thickness of the fibrous cap. Plaque cap thickness of less than about 60 μm is considered to be vulnerable, but this number can vary to some extent depending on the specific patient. The thickness is estimated by using Monte Carlo simulations to determine
30 decorrelation as a function of $r = \sqrt{x^2 + y^2}$ and fitting experimental data to simulations.

C. Mathematical Compensation for Extrinsic Motion

In addition to providing information about the structural features of a plaque, separately analyzing different regions of a speckle pattern also allows decorrelation caused by extrinsic motion to be identified and removed from the analysis. In general, extrinsic motion caused by gross movement of the plaque tissue or blood flow will be directional and global. By contrast, Brownian motion will be non-directional and non-uniform. Thus, by calculating separate decorrelation functions for different regions of the speckle pattern, decorrelation due to extrinsic motion can be identified and subtracted from the functions, allowing isolation of random, Brownian motion. For example, the position of maximums of cross-correlation functions will shift along a vector \vec{v} , which relates extrinsic motion of the sample with respect to the catheter or detection. Brownian motion will decorrelate the speckle patterns in many random directions, and will result in a broadening of the cross-correlation peak and a decrease in correlation maximum above that predicted by linear motion. These two behaviors for intrinsic and extrinsic linear motion should be separable from the cross-correlation function.

V. Additional Imaging Methods

In a simplified system, the rate of decorrelation can be estimated from single pixel speckle images, rather than full, two-dimensional speckle patterns. In this system, a catheter with a single optical fiber could transmit data to a single detector, such as a photodiode. The speckle data gathered would be intensity at the spot as a function of time. From this data, a rate of decorrelation can be calculated directly, without any cross-correlation analysis.

Imaging methods that detect single scattered light, such as optical coherence tomography (OCT) and confocal microscopy, can also be used. While these imaging methods are less sensitive to speckle modulation than the multiple scattering methods described above, they have the advantage of allowing localization of data to a single point within the sample. Such localization would allow measurement of biomechanical properties of the tissue in three dimensions. In addition, in methods that use heterodyne detection, such as OCT, motion of the scatterers can produce a Doppler shift on the returned light. The Doppler shift can provide a further basis for measuring viscosity in the sample. The

mathematics for OCT and confocal microscopy based imaging techniques would be substantially similar to the mathematics described above.

Example

5 In this example, speckle images formed by reflecting laser light from a cadaveric atherosclerotic plaque in a human aorta were analyzed to assess the plaque's viscosity. A portion of normal aorta was also analyzed for comparison.

At a temperature of 37°C, light from a helium-neon laser ($\lambda = 632.8$ nm) was shined on a cadaveric aortic plaque for two seconds. Light reflected from the plaque was received at
10 a CCD camera with a shutter speed of 30 frames per second, through a cross-polarization filter. During the two seconds, the CCD camera recorded a series of 60 speckle images at intervals of 33 ms. Three of the 60 raw data images, corresponding to times $t = 0$, $t = 150$ ms, and $t = 300$ ms, are shown in row A of Fig. 5.

Using IPLab® Spectrum® imaging software, edge detection was performed on the 60
15 raw speckle images, generating 60 edge images. The three edge images for times $t = 0$, $t = 150$ ms, and $t = 300$ ms are shown in row B of Fig. 5. As discussed above, the edge images reflect the spatial derivative of the raw speckle images (i.e., the light patches in the edge images of row B are locations where the intensity is changing as a function of space).

Using the same software, each of the 60 edge images was then compared to the $t = 0$
20 edge image 70 to form 60 cross-correlation images. Each cross-correlation image was generated by multiplying the Fourier transform of the reference image 70 by the complex conjugate of the Fourier transform of the image in question, and then calculating an inverse Fourier transform of the product. For example, referring to row C of Fig. 5, image 72 is an autocorrelation of the $t = 0$ edge image. Image 72 was formed by multiplying the Fourier
25 transform of reference image 70 by the complex conjugate of the Fourier transform of image 70, and then calculating the inverse Fourier transform of the product. Image 74 was formed by multiplying the Fourier transform of image 70 by the complex conjugate of the Fourier transform of the $t = 150$ ms edge image, and then calculating the inverse Fourier transform of the product.

Each cross-correlation image represents the degree of correlation between the corresponding edge image and the reference edge image 70 (i.e., brighter spots are locations where there is a higher degree of correlation than at darker spots).

From each cross-correlation image, the maximum cross-correlation peak (i.e., the correlation at the maximum point of correlation) was calculated using equation (1). The resulting data set included 60 cross-correlation values, each value associated with a time t .

A set of images of normal aorta tissue is shown in Fig. 6. These images are comparable to the set of images in Fig. 5 for a lipid-rich plaque in the same aorta and were imaged and processed in the same manner.

The maximum cross-correlation data for the lipid-rich plaque and the normal aorta tissue were then fit to an exponential cross-correlation function, $G(\tau)$, using Igor Pro®, v 3.01 software (Wavemetrics, Inc). The resulting exponential function was graphed in Fig. 7, curve 80. By way of comparison, the exponential cross-correlation function for speckle data taken from healthy cadaveric aortic tissue is shown in curve 82. The data for curve 82 was gathered and processed using the same procedures as the data for curve 80.

From the cross-correlation data, the decorrelation rate, represented by the time constant τ , was calculated. For the plaque, the time constant was 40 ms. For the aortic tissue, the time constant was 500 ms.

Based on these data, the plaque was borderline vulnerable. Thus, had this plaque been analyzed *in vivo*, using the procedures described above, a physician would have determined that the plaque was a possible candidate for rupture, and may have chosen to intervene, preventing a possible infarction.

OTHER EMBODIMENTS

The methods described herein can also be used to characterize diseased tissue other than atherosclerotic plaques. The microscopic and macroscopic constituents of diseased tissue differ as compared to normal non-pathologic counterparts. For example, speckle patterns can be used to diagnose and characterize other tissue pathology such as neoplasia (cancer), infection, tissue viability, or healing response to injury. In the case of neoplasia, tumors typically have an abnormal abundance of one cell type (clonal) and a surrounding abnormal supporting matrix. This cell type may produce and secrete a viscous fluid, such as

mucin in adenocarcinoma, which would result in lower speckle decorrelation time constants than normal non-cancerous tissue. Moreover, the surrounding matrix may be composed of necrotic tissue and an abundance of abnormal vessels that would also serve to decrease the speckle decorrelation time constant. Other tumors, like osteosarcoma, produce osteoid or
5 immature bone that would increase the time constant compared to normal tissue. Other forms of neoplasia would have increased time constants due to desmoplastic (abundant) fibrous stroma initiated by cytokines produced by the tumor. Indeed, many tumors, including bronchogenic carcinomas and breast carcinomas are firm upon gross examination due to the fibrous stroma surrounding the malignant cells. This fibrous stroma would increase the time
10 constant relative to surrounding normal tissue.

In other examples, in the case of infection, abscesses will be more viscous than surrounding tissue, enabling identification of the infected region by measuring a decrease in the time constant. Inflammation, manifested by the influx of activated inflammatory cells
15 will be characterized by a decrease in the speckle decorrelation time constant as these cells degrade the normal supporting tissue in response to a the presence of bacterial, viral, or foreign body antigens. Necrotic tissue, such as burn eschar, diabetic ulcers, necrotic bowel, and ischemic myocardium will have longer time constants than viable tissue from the same organ due to the lack of intravascular and extravascular fluid and flow in these extracellular spaces.

In the case of healing, fibrosis and fibrous remodeling will likely have longer time
20 constants due to the abundance of collagen matrix and granulation tissue, which would not be present in uninjured tissue. Speckle decorrelation times may also be used to estimate tissue hydration and provide a means for quantifying the state of hydration in a patient. While the above examples elucidate some of the mechanisms that explain how disease affects the
25 biomechanical properties of pathologic tissue, many more exist and are well known in the field of gross anatomic pathology. These differing biomechanical properties can be measured by speckle for the purpose of screening, intraoperative margin identification, and primary diagnosis.

The foregoing detailed description is intended to illustrate and not limit the scope of
30 the invention, which is defined by the appended claims. Other aspects, advantages, and modifications are within the scope of the claims.